

Molecular evolutionary analyses of insect societies

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The social insects live in extraordinarily complex and cohesive societies, where many individuals sacrifice their personal reproduction to become helpers in the colony. Identifying adaptive molecular changes involved in eusocial evolution in insects is important for understanding the mechanisms underlying transitions from solitary to social living, as well as the maintenance and elaboration of social life. Here, we review recent advances made in this area of research in several insect groups: the ants, bees, wasps, and termites. Drawing from whole-genome comparisons, candidate gene approaches, and a genome-scale comparative analysis of protein-coding sequence, we highlight novel insights gained for five major biological processes: chemical signaling, brain development and function, immunity, reproduction, and metabolism and nutrition. Lastly, we make comparisons across these diverse approaches and social insect lineages and discuss potential common themes of eusocial evolution, as well as challenges and prospects for future research in the field.

comparative genomics | molecular evolution | social evolution | sociogenomics

The social insects are exemplars of cooperative group living. Within their complex societies, there is a reproductive division of labor in which only a small number of individuals reproduce, whereas all other individuals belong to a functionally sterile worker caste that specializes in tasks important for colony growth and development (1). Although there has been much theoretical research on the evolutionary forces that may select for eusociality (2, 3), less is known about the actual molecular mechanisms involved in transitions from solitary to social living and in the maintenance and elaboration of eusociality in insects (4).

The social insects provide a powerful comparative framework for investigating mechanisms involved in eusocial evolution. Eusociality has arisen independently at least 12 times in the insects (5–8), and eusocial insects have all converged on the following three characteristics: reproductive division of labor, cooperative brood care, and overlapping generations (9). Additionally, despite sharing this core set of traits, there are many differences among eusocial lifestyles, which may be related to ecological, phylogenetic, or other factors specific to particular eusocial lineages (1). By comparing across social insect lineages, it is possible to both search for common mechanisms of eusocial evolution and explore how eusociality evolves under different conditions.

Analysis of adaptive evolution at the molecular level can yield great insights into the mechanisms underlying the evolution of complex phenotypes, such as eusociality. Genomic sequence provides a molecular record of how natural selection has shaped an organism's evolutionary history (10). Several methods have been developed for comparing genes and genomes to identify molecular signatures of adaptation. These methods were largely developed during the pregenomic era (11) but gain enormous power when large genomic datasets are available, particularly for sets of closely related and phenotypically variable species (12, 13). For example, comparisons of primate genomes have identified adaptive genetic changes involved in the evolution of brain size in humans (14), and comparisons of drosophilid genomes have shed light on the ecological pressures that shaped speciation in this group (13).

Here, we review some of the first contributions of molecular evolutionary research to our understanding of eusocial evolution in insects. This research has focused on the most well-studied social insects, which include several eusocial lineages within the

order Hymenoptera, the ants, bees, and wasps, and the one eusocial lineage in the order Blattodea, the termites (Fig. 1). Some studies have performed targeted molecular evolutionary analyses of candidate genes that have been particularly valuable in species for which large amounts of genomic sequence are not yet available. Others have focused on comparative analyses of whole-genome sequence, which is currently available for six social insects, the honey bee, *Apis mellifera* (15), plus five ant species (16–19), and for many solitary insects, including three solitary hymenopterans in the parasitoid jewel wasp genus, *Nasonia* (20).

We also draw heavily from our own recent genome-scale study of protein-coding sequence evolution in bees (“bee molecular evolution study”). This study analyzed ~3,600 genes from a set of 10 social and nonsocial bee transcriptomes; these species encompass three independent origins of eusociality (21). Hundreds of genes were identified that exhibit a molecular signature of rapid evolution associated with sociality, defined as a higher ratio of non-synonymous-to-synonymous nucleotide substitutions (d_N/d_S) in social relative to nonsocial bee lineages (21). Throughout this review, evidence for rapid evolution is based on relative d_N/d_S and positive selection is defined as $d_N/d_S > 1$, unless otherwise specified.

Genes identified in these studies are listed in Table 1. The insights gained from these studies have implications for understanding how evolutionary changes in the following five major biological processes might be involved in the evolution of eusociality: chemical signaling, brain development and function, immunity, reproduction, and metabolism and nutrition. We discuss evidence and predictions for the putative functional effects of identified molecular changes in these processes on social phenotypes. We also speculate on the potential adaptive significance of these molecular changes and consider whether these changes evolved in response to the origin, maintenance, or elaboration of eusociality, because each case likely involved a distinct set of selective forces. For the purposes of interpreting and synthesizing results across multiple studies, we present each process separately, but it is important to recognize that these biological processes may evolve in concert and that some molecular changes could potentially affect multiple processes. We end with a discussion of future prospects and challenges for this young field.

Chemical Signaling

Social insects use pheromones to coordinate the behavior and physiology of colony members, such as directing the foraging activity of nestmates, reinforcing dominance status, and inhibiting ovary development in workers (45). It is unknown whether chemical signaling was important during the origins of eusociality, because other mechanisms to mediate social interactions, such as physical interactions, serve similar functions in some social insect societies (1). However, chemical signaling is certainly involved in the maintenance and elaboration of eusociality because it is crucial for the coordination and control of colony members. In humans, in whom vocalization is a major compo-

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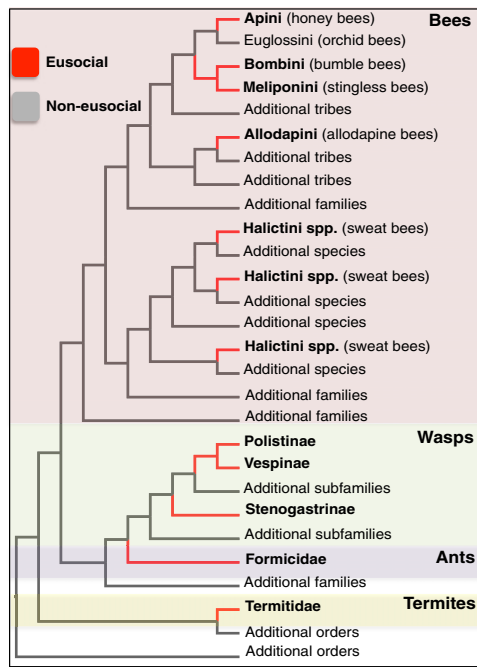


Fig. 1. Cladogram showing the origins of eusociality in insects. Topology and reconstruction of evolutions of eusociality are based on multiple studies (5–8).

nent of social communication, molecular signatures of adaptation have been detected in genes underlying both the production (46) and perception (13) of vocal signals. Early studies in social insects suggest that analogous changes have occurred in the molecular machinery underlying the production and perception of chemical signals.

Gland Development. Our bee molecular evolution study identified ~200 genes evolving more rapidly in social relative to nonsocial bee lineages (21). Gene ontology enrichment analysis revealed that this set of genes was enriched for genes involved in gland development. This supports a role for these genes in chemical signaling, because glands are the primary organs involved in pheromone production in insects. Moreover, the evolution of complex chemical signaling in the social insects has been associated with the diversification of the gland repertoire (1).

In other organisms, modular evolution, in which semiautonomous genetic pathways evolve as a functional unit and are reused in multiple contexts, appears to be a common evolutionary mechanism involved in morphological diversification (47). The sequence changes identified in genes involved in gland development in social bees may have caused modular changes to the gland development program, resulting in functional changes to existing glands or the appearance of entirely new glands. This is supported by the evidence that several of these genes (*decapentaplegic*, *thickveins*, and *PDGF- and VEGF-related factor 1*) have specific roles in gland patterning during early development in *Drosophila* (22, 23).

Because diversification of gland function is a common characteristic shared by all social insects, it would be fruitful to investigate the sequence evolution and function of these genes in other social insect groups. It is possible that molecular changes in the same or similar genes were involved in gland evolution across other independent eusocial lineages.

Odorant Receptors. Given the diversity of chemical signals used by social insects, odorant receptor genes (ORs) have been predicted to be important targets of selection during eusocial evolution (48). Early support for this prediction was found in the genome of the honey bee, *A. mellifera*, which, at the time of its publication, contained the largest number of ORs yet found in an insect genome (15). However, as more insect genomes have been se-

quenced, it has been discovered that *A. mellifera* has an intermediate number of ORs, there is significant variation in OR number between the five ant genomes (16–19), and several solitary insect genomes have among the most ORs found in insects so far (49, 50). Thus, the evidence no longer supports an association between sociality and expansion of the OR repertoire. Furthermore, studies in other organisms have revealed that ORs can function combinatorially and that bioinformatically predicted ORs may not all produce functional proteins, which, together, suggest that the number of ORs in a genome may not scale with the complexity of chemical communication in a species (51).

As a result of their functional specificity, ORs are particularly good targets for candidate gene studies, because the adaptive significance of OR evolution may be easier to interpret than for genes with broader functions (51). A functional genomics approach was used to identify a novel OR in the *A. mellifera* genome, *AmOr11*, which responds to the main component of the honey bee queen pheromone, (*E*)-9-oxo-2-decenoic acid (9-ODA) (24). The queen pheromone attracts workers to the queen, partially inhibits worker ovary development, and acts as a sex pheromone, among other functions (24). The specific molecular characteristics of *AmOr11* that are involved in the perception of 9-ODA are not yet known, but it appears that it arose early in *Apis* evolution (52–54).

Termite Queen Pheromone. *Neofem2* is the first gene discovered in termites that is involved in signaling queen presence to workers. It was originally identified as being up-regulated in female neotenic “replacement” reproductives relative to other colony members in two species of *Cryptotermes* termites (25). Knocking down *Neofem2* in *Cryptotermes secundus* queens using RNAi caused an increase in aggressive behavior among workers, which is typically only exhibited under queenless conditions (25). Based on sequence similarity, *Neofem2* is most closely related to a β -glycosidase expressed in the salivary glands of the termite *Neotermes koshunensis* (26). β -glycosidases are enzymes that break down polysaccharides; in wood-dwelling termites, such as *N. koshunensis* and *C. secundus*, whose diet primarily consists of rotting bark, these enzymes are important for breaking down cellulose (55). It has thus been suggested that *Neofem2* evolved from a wood-digesting enzyme to pheromone (26). Supporting this speculation, β -glycosidases exhibit pheromonal activity in other insects, including the production of an egg recognition signal in another termite species (26). The specific molecular changes that have occurred in *Neofem2* as it evolved this new social function remain to be discovered. The story of *Neofem2* highlights the importance of considering the ecological context of social evolution in a given lineage, because the origin of a social pheromone from a wood-digesting enzyme is almost certainly a phenomenon specific to the wood-dwelling termites.

General protein-9 in Fire Ants. *General protein-9* (*Gp-9*) alleles are strongly associated with variation in queen number in fire ants (genus *Solenopsis*). In monogynous (single queen) colonies, all females are homozygous for *B*-type alleles and will not tolerate the presence of multiple queens, whereas in polygynous (multiple queens) colonies, some individuals possess *b*-type alleles and do accept multiple queens but only if those queens also possess the *b*-type allele (27). *Gp-9* has been called a “green beard gene” (28), because workers carrying one allele favor queens that share the same allele. Molecular phylogenetic analyses of *Gp-9* both within and across *Solenopsis* species have revealed that the *b*-like alleles form a monophyletic clade, suggesting that monogyny was the ancestral condition in the genus and that polygyny arose once and has been maintained through multiple speciation events (29, 30).

At the protein sequence level, *Gp-9* most closely resembles odorant-binding proteins (OBPs), which are expressed in chemosensory sensilla lymph and bind and transport soluble odorants (27). These results have led to the suggestion that *Gp-9* is an OBP that plays a role in pheromonal communication in fire ants (27). However, *Gp-9* is ubiquitously expressed in the hemolymph, suggesting it may be involved in functions that are unrelated to chemosensation (31). In addition, *Gp-9* is found in a genomic region with a low recombination rate; therefore, other linked genes in the

Table 1. Genes implicated in the origin or maintenance of insect society by molecular evolutionary research

Gene	Function	Evidence	Type of change*
Chemical signaling			
<i>decapentaplegic</i>	Gland development (22, 23)	Rapid evolution in eusocial bees (21)	1
<i>thickveins</i>	Gland development (22, 23)	Rapid evolution in eusocial bees (21)	1
<i>PDGF- and VEGF-related factor 1</i>	Gland development (22, 23)	Rapid evolution in eusocial bees (21)	1
<i>AmOr11</i>	OR (24)	Responds to main component of queen honey bee pheromone, 9-ODA (24)	2
<i>Neofem 2</i>	β-Glycosidase-like (25, 26)	Involved in signaling queen termite presence (25, 26)	3
<i>GP-9</i>	Putative OBP (27–31)	Allelic variation associated with fire ant queen number (27–31)	1, 2
Brain development and function			
<i>dunce</i>	cAMP/CREB signaling pathways (32)	Rapid evolution in primitively eusocial bees (21)	1
<i>nejire</i>	CREB binding protein (32)	Rapid evolution in primitively eusocial bees (21)	1
Immunity			
<i>defensin</i>	Antimicrobial protein (33)	Positive selection in ants (33)	1
<i>termicin</i>	Antimicrobial protein (34, 35)	Gene duplication, positive selection in termites (34, 35)	1, 2
<i>GNBP 1 and 2</i>	Pattern recognition receptors (36)	Gene duplication, positive selection in termites (36)	1, 2
<i>relish</i>	Transcription factor, induces production of antimicrobial peptides (36)	Positive selection in termites (36)	1
Reproduction			
<i>tudor</i>	piRNA pathway (37)	Rapid evolution in primitively eusocial bees (21)	1
<i>capsuleen</i>	piRNA pathway (37)	Rapid evolution in primitively eusocial bees (21)	1
<i>vasa</i>	piRNA pathway (37)	Rapid evolution in primitively eusocial bees (21)	1
<i>csd</i>	Sex determination (38–40)	Gene duplication, positive selection in honey bees (38–40)	1, 2
Metabolism and nutrition			
<i>MRJPs</i>	Main components of royal jelly (41)	Gene family expansion, novel feeding-related functions in honey bees (41)	2
<i>Hex-1 and Hex-2</i>	Storage proteins (42, 43)	Unique insertions in termites (42, 43)	1
<i>phosphofructokinase</i>	Key regulator of glycolysis (44)	Rapid evolution in eusocial bees (21)	1
<i>hexokinase</i>	Regulator of glycolytic flux (44)	Rapid evolution in eusocial bees (21)	1
<i>pyruvate kinase</i>	Regulator of glycolytic flux (44)	Rapid evolution in eusocial bees (21)	1

Although many genes in this table are presumably involved in multiple biological processes, they are classified in one of five processes with known links to insect sociality: chemical signaling, brain development and function, immunity, reproduction, and metabolism and nutrition.

*Type of change: 1, protein coding sequence change; 2, novel gene; 3, change unknown.

region may potentially have more influence on the regulation of queen number (27, 29). *Gp-9* alleles are also associated with variation in several life history traits in *Solenopsis* queens, including body fat and dispersal behavior (30), suggesting that *Gp-9* either acts pleiotropically or with other genes in the region.

Although the function of *Gp-9* is unresolved, molecular evolutionary analyses suggest that this gene is evolving adaptively, implying that *Gp-9* played an important role in fire ant evolution. A signature of positive selection was detected in the branch leading to the *b*-like allele clade (29), suggesting that this allele had an adaptive benefit when it arose. In addition, all *b*-like alleles share the same amino acid residues at three diagnostic codon positions, and two of these positions show evidence of positive selection in *Solenopsis invicta*, the species in which it has been best studied (30).

Brain Development and Function

Some of the most striking differences between social and solitary insects are behavioral. Several social insect behaviors appear to be truly novel, such as symbolic dance communication in honey bees and slave making in ants (1). Other behaviors exhibited by social insects appear to be modified forms of behaviors performed by solitary insects, for example, social foraging, which resembles nest provisioning in solitary insects. It is likely that molecular changes affecting nervous system development and

function were important in the evolution of social insect behaviors, but very little is currently known.

Brain Evolution in Primitively Eusocial Bees. Our bee molecular evolution study detected a strong signal of rapid evolution in brain-related genes in primitively eusocial, but not highly eusocial lineages across two independent origins of each lifestyle (21). Among these rapidly evolving genes were *dunce* and *nejire*, two genes that mediate learning and memory in invertebrates and vertebrates through cAMP/CREB signaling pathways (32).

The detection of molecular changes in brain-related genes exclusively in primitively eusocial bee lineages is perhaps surprising, given that this finding is not what may have been predicted by a prominent hypothesis about the relationship between sociality and brain evolution in vertebrates, the social brain hypothesis (SBH). Originally developed to explain the evolution of the enlarged neocortex in many social vertebrates, the SBH posits that the cognitive demands of social living are a strong selective force in brain evolution (56). Given that highly eusocial bee societies have larger colony sizes, greater social complexity, and novel behaviors (i.e., dance communication in honey bees) relative to primitively eusocial bees, one might have assumed that the cognitive demands of social living are strongest in highly eusocial species and lead to stronger selection on brain-related genes.

Unique features of insect sociality and the primitively eusocial lifestyle may help to explain why selection on brain evolution appears to have been stronger in the primitively eusocial bees.

First, unlike in vertebrate social evolution, where there has been an emphasis on increased individual cognitive abilities, there appears to have been an emphasis on increased connectedness among colony members in insect social evolution, often accompanied by a reduction of individual behavioral repertoires (57, 58). Therefore, individual cognitive abilities may not be correlated with group size in social insects, as has been found in vertebrates. There are also several distinguishing features of the primitively eusocial bee lifestyle that may have placed unique selective pressure on brain evolution in these lineages. Social structure in primitively eusocial bee colonies is typically maintained through fluid and dynamic dominance hierarchies (9, 59), which can be an especially cognitively challenging form of social interaction (59, 60). In addition, a primitively eusocial bee queen is capable of behaving both solitarily, as she does during the colony-founding phase of her lifecycle, and socially, as she does once she has reared her first brood of workers (9).

In both ants and wasps, which each evolved eusociality independent of bees, there are some species in which queens exhibit a similar “solitary-like” phase during colony founding and other species that found colonies in swarms, like highly eusocial bees do (1). A comparison of brain-related genes and/or brain structure in ant and wasp species that do or do not establish colonies solitarily may provide clues as to whether this trait is a strong force in social insect brain evolution. One study in paper wasps reported brain region volume differences between swarm and independent-founding species, suggesting that these differences in colony founding can affect brain evolution (61).

Immunity

Pathogens and parasites are thought to have been a strong selective force challenging the maintenance of sociality in a variety of organisms, including social insects (62). Crowded living conditions, often with closely related individuals, facilitate pathogen transmission (62). Social insects appear to have responded to this potentially dissolutive selective pressure in three main ways (33). The first way is through “social immunity,” which refers to group-level defenses, such as hygienic behaviors and the use of collected antimicrobial resins for lining nest cavities (62). The second way is through increasing intracolony genetic diversity via multiple mating by queens (63) and high rates of genetic recombination (4) to enhance colony-level disease resistance. The third way is through adaptive evolution of immune genes (33).

Molecular evolutionary analyses of immune genes have provided some of the best examples of positive selection acting in social insect genomes. This may be partly attributable to the fact that immune systems, in general, are often at the forefront of an ongoing evolutionary arms race with pathogens; thus, selection pressure on immune-related genes is typically quite strong (64). In addition, many immune-related genes are functionally well-characterized (65), facilitating interpretations of the adaptive significance of sequence changes.

Immune Gene Evolution in Hymenoptera. When the first social insect genome was sequenced, that of *A. mellifera*, researchers were intrigued by the low number of immune genes found in *A. mellifera* relative to other fully sequenced insect genomes: those of the Diptera, *Drosophila melanogaster* and *Anopheles gambiae* (15). Although the main components of canonical immune pathways are conserved, the *A. mellifera* genome contains smaller numbers of gene family members at all points along these pathways (66). It was hypothesized that the loss of immune genes was facilitated by novel forms of social immunity in social insects, resulting in relaxed constraint on immune genes (66). However, as more insect genomes have been sequenced, it has become apparent that sociality is not necessarily predictive of immune gene number. Rather, it seems that dipterans have unusually large immune gene repertoires, whereas the recently sequenced ant genomes (16–19); the solitary wasps, *Nasonia* (20); and the solitary pea aphid, *Acyrtosiphon pisum* (67) have similar numbers of immune genes as *A. mellifera* (66).

By contrast, molecular evolutionary analysis of individual immune genes in social Hymenoptera has provided evidence that sociality has driven immune gene sequence evolution. One study

revealed that some immune genes are evolving more rapidly in species of honey bees, bumble bees, and ants relative to *Drosophila* (68). This study also showed that immune genes are evolving more rapidly than nonimmune genes in several honey bee species. Similarly, genes related to innate immunity and humoral immunity were among the fastest evolving (based on branch lengths in phylogenetic trees inferred from protein sequence) in *A. mellifera* in a comparison of over 3,000 genes among *A. mellifera*, *Nasonia*, and their common ancestor (20). Additionally, evidence for positive selection has been detected in the antimicrobial protein *defensin* in a study comparing the sequence of 27 ant species (33). This study revealed that the signal and propeptide regions of *defensin*, which are cleaved off to activate the mature peptide, are evolving neutrally, whereas the active region of the peptide is under positive selection, including one amino acid site thought to mediate antimicrobial activity. Our bee molecular evolution study did not detect a strong signal of selection on immune genes, but that was likely because these classes of genes were underrepresented in our dataset (21).

Immune Gene Evolution in Termites. A study of the termite defensin-like gene, *termicin*, in 11 *Nasutitermes* termite species revealed that this gene has duplicated repeatedly during *Nasutitermes* radiation and that positive selection has driven a divergence in the molecular charge of the gene copies (34). Insect defensins are known to function by disrupting bacterial plasma membranes, and experimental evidence suggests that molecular charge may be a crucial component of this activity (34). It was hypothesized that there is a selective advantage to having two termicins with different charge properties at specific sites (34). In support of this hypothesis, results from this study suggest that ancestral termicins had relatively high positive charges and that in species in which there has been a gene duplication event, positive selection has driven a decrease in charge for one of the copies. Sequence analysis revealed a strong positive correlation between the strength of selection (d_N/d_S) and the change in molecular charge along different termicin lineages. Additionally, three amino acid sites that show a signature of positive selection have substitutions at these sites that contribute to a charge change, and they fall on the external surface of the predicted protein structure, suggesting that these sites may interact with a fungal membrane receptor (34).

A different study of 13 *Nasutitermes* termite species also found evidence that gene duplication and positive selection are involved in termite immune gene evolution (36). This study focused on genes encoding Gram-negative bacterial-binding protein 1 and 2 (GNBP1 and GNBP2), which are thought to have duplicated early in termite evolution, and the transcription factor *relish*, which induces production of antimicrobial peptides in *Drosophila*. All three genes show evidence of positive selection, with *relish* showing the strongest signal. Four of the five positively selected sites in *relish* are in a “spacer” region of the protein that is cleaved by the caspase Dredd. This cleavage is thought to activate *relish* by generating a DNA-binding Rel homology domain that translocates to the nucleus and binds to promoters of target genes (69). Analysis of the *Drosophila simulans* ortholog also found positive selection in this spacer region (36). It was hypothesized that microbial pathogens may be targeting this region of *relish* to prevent its activation, sparking an evolutionary arms race as *relish* evolves counterresponses to maintain its normal function (36). Another study found evidence of positive selection in *termicin* but not in GNBP2 in two *Reticulitermes* termite species, a genus distantly related to the *Nasutitermes* genus (35). This study used a population genetics approach to analyze intraspecific polymorphism and interspecific divergence in coding sequence, and results indicated that *termicin* underwent a selective sweep driven by positive selection for beneficial amino acid changes.

Reproduction

In many insect societies, queens are highly reproductive individuals, whereas workers perform almost no reproduction activity. Worker sterility is achieved through a variety of morphological, behavioral, and physiological mechanisms in social insects (1). For example, in many social species, workers lack spermatheca for

sperm storage. In addition, ovary development is tightly regulated by social cues, and queens and workers typically have grossly over- and underdeveloped ovaries, respectively, relative to solitary insects (1). Sociality also has strong implications for reproductive behavior, particularly for mating frequency, which can affect genetic variation among colony members.

Ovary Development in Primitively Eusocial Bees. Our bee molecular evolution study identified some genes involved in ovary development evolving most rapidly in primitively eusocial bees (21). Although both highly and primitively eusocial bee societies have a strong reproductive division of labor, the reproductive differences between queen and worker in primitively eusocial species are less extreme, and ovary development appears to be more sensitive to social cues in primitively eusocial species (1). Perhaps the molecular changes in ovary development-related genes found only in the primitively eusocial lineages underlie some of the unique characteristics of the reproductive biology of this eusocial lifestyle.

Several genes (i.e., *tudor*, *capsuleen*, *vasa*) evolving rapidly in one or both of the primitively eusocial bee lineages interact together in the PIWI RNA (piRNA) pathway. The piRNA pathway is expressed only in gametic tissue, and it is involved in regulating gametic cell division and differentiation (37). Functional PIWI genes have recently been discovered in *A. mellifera* (70), suggesting that the piRNA pathway is present and functional in bees. These genes are particularly good candidates for further study, because the tissue specificity of the piRNA pathway suggests that selection on these genes is specifically directed at changes related to reproductive processes, in contrast to genes with broader ranges of tissue expression, where the functional target of selection is harder to infer. Additional ovary development-related genes unrelated to the piRNA pathway also showed a signature of rapid evolution in these primitively eusocial bees (21).

Sex Determination and complementary sex determiner in Honey Bees. More is known about the evolution of *complementary sex determiner* (*csd*) in honey bees than probably any other gene in the social insects. The story of *csd* involves the origin of entirely new genes and pathways, as well as a classic example of balancing selection. Sex determination in honey bees is based on genotype at the *csd* locus; individuals heterozygous at the *csd* locus develop into females, whereas hemizygous individuals develop into males (38). Sex determination in many Hymenoptera is probably determined through a similar single-locus system of complementary sex determination (71), but *csd* is the first and only locus that has been discovered thus far. The genomic region containing *csd* was first identified through mapping (38), and the function of the gene was confirmed by RNAi, which showed that reducing *csd* expression in genetically female eggs results in male-like development (40). Complementary sex determination not only regulates sex determination but influences many aspects of social insect biology that are influenced by kinship and degrees of relatedness, including kin selection and the genetic composition of colonies, which are important for division of labor and colony immunity (4).

csd appears to be a honey bee-specific gene because it has been found in multiple *Apis* species (39) but not outside of the genus (40). The gene likely evolved through the duplication of an adjacent gene, *feminizer* (*fem*). *csd* and *fem* are similar (>70%) in amino acid sequence, and both are serine/arginine-rich proteins, a class of proteins involved in RNA splicing (40). Both genes share two major domains, but *csd* has an additional hyper-variable region located between these other domains (40). *fem* has been found in several non-honey bee species and in *Nasonia* wasps, but not in any additional insect species, suggesting that it evolved sometime before the split between the hymenopteran superfamilies Apoidea and Chalcidoidea ~140 Mya but after the split from *Drosophila* ~300 Mya (40). *fem* shares some functional and sequence similarities to *transformer* (*tra*), a gene involved in sex determination in *Drosophila*, and it perhaps evolved from an ancestral form of *tra* common to fly and bee lineages (38, 40). RNAi experiments were used to show that *csd* acts upstream of *fem* in the sex determination pathway. Genetically female embryos treated with *fem* RNAi develop male heads, and RNAi

knockdowns of *csd* cause male-specific *fem* splicing, suggesting that *csd* is involved in *fem* splicing (40).

csd has been subject to rigorous population genetic analysis. Because homozygous males do not reproduce, it was predicted that there would be strong negative frequency-dependent selection at the *csd* locus (39). This prediction has been upheld, because at least 15 different *csd* alleles have been found in natural populations around the world in three different *Apis* species (39) and the gene has accumulated 10- to 13-fold more mutations than the rest of the genome (39). Pairwise nonsynonymous differences between alleles are highest in exons 6 and 7 (39), suggesting that this region is a target of positive selection, and is therefore presumably functionally important. Six fixed amino acid differences between *csd* and *fem* are located in the coiled-coil domain, which is important in protein binding (40). Strong positive selection was detected on the branch right after the split between the two genes, suggesting that positive selection played a role in their diversification (40).

Metabolism and Nutrition

Transcriptomic analyses have shown that nutritional and metabolic pathways play an important role in queen-worker caste determination in every eusocial insect lineage thus far studied and also contribute to worker-worker division of labor in many species (4). Given these fundamental connections to eusociality, nutritional and metabolic pathways are well-studied in social insects and several molecular evolutionary studies have identified changes associated with their function.

Major Royal Jelly Proteins. The evolution of the Major Royal Jelly Proteins (MRJPs) in honey bees is an excellent example of novel genes playing an integral role in the social biology of a species. In the honey bee, *A. mellifera*, the developmental fate of female larvae is determined by the amount of royal jelly they consume (72). Royal jelly is a protein- and lipid-rich substance secreted from the hypopharyngeal glands of brood-feeding “nurse” bees and fed to larvae, which triggers endocrine and epigenetic events that lead to the development of either a worker or a queen (72, 73). The main components of royal jelly are the MRJPs. The *A. mellifera* genome contains 10 *mrjp* genes, encoding 9 MRJPs (one *mrjp* is a pseudogene). These genes are arranged in tandem in the genome, have high sequence similarity (~60%) to one another, and have a conserved intron/exon structure, suggesting that they are a fairly young gene family (41). There is evidence that *mrjp* genes are also present in other *Apis* species (41, 74).

The *mrjp* gene family in *A. mellifera* appears to have evolved via a gene duplication event from a member of the *yellow* gene family. The cluster of *mrjp* genes in the *A. mellifera* genome is flanked by members of the *yellow* gene family, and one of the flanking *yellow* genes, *yellow-e3*, shares the characteristic intron/exon structure of the *mrjp* genes, suggesting that it is their progenitor (41). Members of the *yellow* gene family are involved in pigmentation, reproductive physiology, and courtship behavior in insects (75).

The use of *mrjp* genes for larval feeding appears to be a derived social trait that is unique to honey bees. Although *mrjp*-like genes have been found in other social and nonsocial Hymenoptera species, evidence suggests that the *yellow* gene family is prone to duplication and that the *mrjp*-like genes in non-*Apis* species evolved independent of *Apis* (19, 20). Furthermore, there is no evidence of a food-related role for any *mrjp*-like or *yellow*-like gene outside of *Apis* (75). Because many other social insect species manipulate larval nutrition for the purposes of caste determination without the use of specialized glandular secretions (76), the evolution of the *mrjp* genes in honey bees appears to be associated with the elaboration of eusociality and may have been correlated with or dependent on other evolutionary changes, such as changes in gland function.

Hexamerins. The work done on the termite hexamerins is another excellent example of linking genetic changes to protein function and social phenotype. In the lower termites, workers may develop into either reproductives or soldiers, depending on a number of social and environmental cues, and differentiation into the sol-

dier caste is induced by high juvenile hormone (JH) titers (42). RNAi studies in the termite *Reticulitermes flavipes* have shown that two hexamerin genes, *Hex-1* and *Hex-2*, are involved in the regulation of this caste determination (43). In many insects, hexamerins act as storage proteins that sequester substances from the diet and release them when food is scarce or inaccessible, such as during early development (42). It has been hypothesized that *Hex-1* and *Hex-2* work together to regulate caste differentiation in termites via direct interactions with JH (43); however, elucidating the specific molecular mechanisms involved in JH action is a difficult challenge in insects in general (77).

Molecular evolutionary studies of *Hex-1* and *Hex-2* provide clues as to how these genes may interact with JH. Relative to 100+ known *Hex* genes in other insects, both termite *Hex* genes have distinctive insertions in their coding regions; the unique insertion in *Hex-1* contains a prenylation motif with a proposed function in JH binding, and the unique insertion in *Hex-2* shares sequence similarities to the well-characterized blowfly (*Calliphora vicina*) hexamerin receptor (43). Consistent with these predicted functions, follow-up experiments demonstrated that the *Hex-1* protein has strong binding affinity for JH and the *Hex-2* protein shows strong membrane affinity, as would be expected for a receptor protein (43).

Hexamerins also exhibit novel social functions in other social insect species, suggesting that they may be particularly prone to social co-option. Evidence in honey bees (78) and *Polistes* wasps (79) suggests that hexamerins may be important in caste determination in these social insect lineages, and in ants, hexamerins appear to have been important in the evolution of elaborated life history characteristics (80).

JH, Insulin, and Vitellogenin Axis. In the highly eusocial honey bee, *A. mellifera*, the JH and insulin/insulin-like growth factor-1 (IIS) signaling pathways, as well as the yolk protein precursor *vitellogenin* (*vg*), interact with one another and function in novel ways that are important in multiple social contexts. JH does not function as a gonadotropin in adult honey bees as it does in most insects; instead, it plays a strong role in caste determination and worker division of labor (81). The IIS signaling pathway interacts with JH and is also involved in worker division of labor. Foragers exhibit higher expression of genes in the IIS pathway in the brain relative to nurses, and down-regulating IIS signaling delays the age-related transition from nursing to foraging (82). This represents a reversal of the traditional positive relationship between high nutrition and IIS signaling, because foragers are nutritionally deprived relative to nurses (82). *Vg* also shows novel social functions in honey bees. It is highly expressed in some workers, although they are largely nonreproductive; it may be used by nurses in the synthesis of royal jelly (83); and it functions as an antioxidant that may be involved in promoting longevity in queen bees (84).

The molecular changes underlying these novel functions of JH, IIS, and *Vg* are unknown, but insights from solitary insects may provide clues as to what these changes may be. The relationship between genetic variation and regulation of JH titers has been particularly well-studied in crickets and butterflies (85), molecular evolution and function of the IIS pathway have been investigated across the complete genomes of 12 *Drosophila* species (86, 87), and insect *Vgs* and their receptors are well-characterized at the molecular level (88).

Carbohydrate Metabolism. Several studies in bees suggest that the evolution of the highly eusocial lifestyle involved molecular changes in genes related to carbohydrate metabolism. Our bee molecular evolution study revealed that genes involved in carbohydrate metabolism are evolving more rapidly in eusocial relative to noneusocial bee lineages and are evolving most rapidly in highly eusocial lineages (21). In particular, 15 genes encoding glycolytic enzymes showed evidence of rapid evolution in eusocial lineages, including enzymes that play a key regulatory role (e.g., *phosphofructokinase*) or are involved in glycolytic flux (e.g., *hexokinase*, *pyruvate kinase*) (44). Analysis of protein sequence evolution of genes with queen-biased brain gene expression in *A. mellifera* found that queen-biased genes involved in metabolism, including carbohydrate metabolism, were among

the most rapidly evolving (based on branch lengths in phylogenetic trees inferred from protein sequence) relative to orthologs from several solitary insects (89). Comparative analysis of the genome sequences of *A. mellifera*, *D. melanogaster*, and *A. gambiae* suggest that there may also have been bee-specific changes in gene copy number for carbohydrate-metabolizing genes (44). Given that carbohydrate metabolism is such a fundamental “house-keeping” process, it is not immediately clear why there has been unique selective pressure on these processes in highly eusocial bee lineages. Here, we offer three speculative hypotheses.

First, increases in the flight demands of highly eusocial bees may have placed strong selective pressure on increasing efficiency of glycolytic enzymes, because carbohydrates are the main fuel for flight in bees (90). The individual foraging activity of highly eusocial bee workers appears to be higher than for solitary bees (91), although, to the best of our knowledge, no direct comparisons of highly and primitively eusocial bee foraging activity have been performed.

Second, highly eusocial bees are unique in relying exclusively on a diet of modified stored sugars (i.e., honey) for long periods of time. Nest thermoregulation during winter months is completely reliant on honey stores as a fuel source to sustain workers, who shiver to produce metabolic heat to maintain optimal hive temperature (92). Perhaps these differences in diet have placed some novel selective pressure on glycolytic enzymes in highly eusocial lineages.

Third, perhaps the greatly extended life span of queens in highly eusocial species evolved through changes in metabolism-related genes, including those involved in carbohydrate metabolism. A connection between reduced metabolic rate and increased life span has been shown in many species (93). In the honey bee, *A. mellifera*, queens exhibit an age-related reduction in IIS signaling (84) that regulates carbohydrate metabolism. If the molecular changes in carbohydrate metabolism genes in highly eusocial bees were attributable to selection for extended queen life span, it can be predicted that similar molecular changes may also be found in independent social insect lineages that also exhibit extended queen life spans (1).

Prospects and Challenges

Recent work on molecular evolutionary changes in social insects has identified specific genes, molecular pathways, and biological processes that appear to have been shaped by natural selection. Some of these changes can be plausibly associated with the origins, maintenance, or elaboration of eusociality, albeit speculatively.

Two insights emerge from this review. First, it appears that there have been unique genetic changes in different social insect lineages, suggesting that the multiple independent occurrences of eusociality have involved multiple molecular routes. These differences may reflect distinct ecological or other constraints for each lineage. For example, the evolution of a queen pheromone in termites from a wood-digesting enzyme seems fitting, given that many termite societies live in rotting wood (26).

Second, genetic changes also have occurred in similar biological functions across diverse species of social insects. This supports the concept of a genetic toolkit for eusociality (94). This concept is reasonable, because despite the striking diversity among social insect species, they all have converged on a similar suite of traits, which are the defining characteristics of eusociality (9). Previous research suggesting components of a genetic toolkit for eusociality has focused on genes and molecular pathways that are associated both with solitary and related social behaviors in insects, for example, the *foraging* gene, which is involved in feeding behavior in *Drosophila* and a variety of other solitary organisms, and social foraging behavior in honey bees and ants (94). Transcriptomic studies have also identified shared sets of genes whose expression patterns are associated with division of labor in independent social insect lineages (95).

The molecular evolutionary studies we reviewed identify biological processes and specific genes that may be excellent systems in which to investigate the concept of a genetic toolkit for eusociality further. Among the most promising are the following:

- i) *Hexamerins*. As discussed above, hexamerins have been shown to be involved in queen physiology and other social traits in a variety of social insects, and the work on *Hex-1* and *Hex-2* in termites demonstrates how hexamerin sequence evolution can be studied and linked to social traits.
- ii) *Gland development genes*. The rapidly evolving gland development genes identified in our bee molecular evolution study (21) are also good candidates for further study, because the gene functions are relatively well-characterized and gland diversification is a universal phenomenon in social insect evolution.
- iii) *Brain-related genes*. The rapidly evolving brain-related genes identified in primitively eusocial lineages in our bee molecular evolution study (21) are prime candidates for further study in primitively eusocial bees, as well as in ant and wasp species that share the primitively eusocial bee lifestyle feature of solitary nest-founding.

The molecular changes and biological processes highlighted in this review are currently the most well-studied in social insects. There are almost certainly other equally important types of molecular changes and biological processes associated with social insect evolution that have not yet been discovered, perhaps because of the limited range of taxa subjected to these type of analyses thus far. This gap in our knowledge is largely attributable to a lack of genomic resources, especially for closely related social and nonsocial species. For example, some types of genetic changes, such as chromosomal rearrangements and patterns of DNA methylation, are not possible to study with only fragments of the genome. In addition, the identification of truly novel genes is limited by the small sample size of available genomes and less well-developed forward genetic analyses in social insects relative to model genetic organisms. As these limitations are overcome, it should be possible to search more broadly for different types of genetic changes associated with the evolution of eusocial traits. These analyses can be guided by several theoretical models that have been proposed to predict the types of genetic changes that are most important in social evolution (96–98).

Whole-genome scans for molecular signatures of adaptive evolution specific to social insects will be particularly useful for generating new hypotheses and implicating new biological processes in social insect evolution. Candidate gene approaches across a broad sample of social and nonsocial insects will allow for greater accuracy in reconstructing the phylogenetic history of molecular changes and testing their associations with social

evolution. Once specific sequence changes are identified, functional analyses are necessary to determine their effect on protein-, organismal-, and group-level phenotypes, as well as the adaptive significance of the phenotype change (99).

This leads us to raise one important caveat for most molecular evolutionary studies in the social insects: the lack of species-specific information about gene function. As is often the case, gene function in this paper is typically inferred from orthology to the fruit fly *D. melanogaster*, which shared a common ancestor with eusocial insect lineages over 300 Mya (15). Although gene function for molecular processes is generally highly conserved over evolutionary time, when interpreting findings, it is important to consider the possibility that a particular gene has evolved a novel function. Furthermore, many genes have multiple functions; thus, the target of selection can be difficult to infer solely from identifying molecular evolutionary changes. Experimental approaches to determining gene function in social insects, via RNAi and transgenesis, will strengthen the interpretation of molecular evolutionary findings. Additional challenges arise in determining the adaptive or ecological significance of molecular changes, even when their functional significance is understood (100).

Despite these challenges, molecular evolutionary analysis of social insect societies holds promise for testing venerable theories of social evolution using genomic data. Multiple evolutionary scenarios have been proposed as potential routes to group living in insects. These include the composition of incipient social groups, such as associations between mothers and offspring (the “subsocial” route) or between related and unrelated individuals of the same generation (“semisocial” route) (9); mechanisms through which altruism is achieved, such as kin selection (2); parental manipulation of offspring or voluntary helpers at the nest (101); and necessary preadaptations for social living, such as a monogamous mating system (102) or progressive provisioning of offspring (3). Wedding this rich theory with genome-scale molecular evolutionary analysis and functional experimentation holds the promise of finally answering the compelling question of how eusociality evolved in insects.

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1. Wilson EO (1971) *The Insect Societies* (Belknap Press, Cambridge, MA).
2. Strassmann JE, Queller DC (2007) Insect societies as divided organisms: The complexities of purpose and cross-purpose. *Proc Natl Acad Sci USA* 104(Suppl1): 8619–8626.
3. Nowak MA, Tarnita CE, Wilson EO (2010) The evolution of eusociality. *Nature* 466: 1057–1062.
4. Smith CR, Toth AL, Suarez AV, Robinson GE (2008) Genetic and genomic analyses of the division of labour in insect societies. *Nat Rev Genet* 9:735–748.
5. Cameron SA, Mardulyn P (2001) Multiple molecular data sets suggest independent origins of highly eusocial behavior in bees (Hymenoptera:Apinae). *Syst Biol* 50:194–214.
6. Cardinal S, Straka J, Danforth BN (2010) Comprehensive phylogeny of apid bees reveals the evolutionary origins and antiquity of cleptoparasitism. *Proc Natl Acad Sci USA* 107:16207–16211.
7. Hines HM, Hunt JH, O'Connor TK, Gillespie JJ, Cameron SA (2007) Multigene phylogeny reveals eusociality evolved twice in vespid wasps. *Proc Natl Acad Sci USA* 104:3295–3299.
8. Brady SG, Sipes S, Pearson A, Danforth BN (2006) Recent and simultaneous origins of eusociality in halictid bees. *Proc Biol Sci* 273:1643–1649.
9. Michener CD (1974) *The Social Behavior of Bees* (Belknap Press, Cambridge, MA).
10. Clark AG (2006) Genomics of the evolutionary process. *Trends Ecol Evol* 21:316–321.
11. Li WH (1997) *Molecular Evolution* (Sinauer Associates, Sunderland, MA).
12. *Drosophila* 12 Genomes Consortium (2007) Evolution of genes and genomes on the *Drosophila* phylogeny. *Nature* 450:203–218.
13. Clark AG, et al. (2003) Inferring nonneutral evolution from human-chimp-mouse orthologous gene trios. *Science* 302:1960–1963.
14. Pollard KS, et al. (2006) An RNA gene expressed during cortical development evolved rapidly in humans. *Nature* 443:167–172.
15. Honeybee Genome Sequencing Consortium (2006) Insights into social insects from the genome of the honeybee *Apis mellifera*. *Nature* 443:931–949.
16. Bonasio R, et al. (2010) Genomic comparison of the ants *Camponotus floridanus* and *Harpegnathos saltator*. *Science* 329:1068–1071.
17. Wurm Y, et al. (2011) The genome of the fire ant *Solenopsis invicta*. *Proc Natl Acad Sci USA* 108:5679–5684.
18. Smith CR, et al. (2011) Draft genome of the red harvester ant *Pogonomyrmex barbatus*. *Proc Natl Acad Sci USA* 108:5667–5672.
19. Smith CD, et al. (2011) Draft genome of the globally widespread and invasive Argentine ant (*Linepithema humile*). *Proc Natl Acad Sci USA* 108:5673–5678.
20. Werren JH, et al.; Nasonia Genome Working Group (2010) Functional and evolutionary insights from the genomes of three parasitoid *Nasonia* species. *Science* 327:343–348.
21. Woodard SH, et al. (2011) Genes involved in convergent evolution of eusociality in bees. *Proc Natl Acad Sci USA* 108:7472–7477.
22. Bradley PL, Myat MM, Comeaux CA, Andrew DJ (2003) Posterior migration of the salivary gland requires an intact visceral mesoderm and integrin function. *Dev Biol* 257:249–262.
23. Harris KE, Schnitke N, Beckendorf SK (2007) Two ligands signal through the *Drosophila* PDGFR/VEGF receptor to ensure proper salivary gland positioning. *Mech Dev* 124:441–448.
24. Wanner KW, et al. (2007) A honey bee odorant receptor for the queen substance 9-oxo-2-decenoic acid. *Proc Natl Acad Sci USA* 104:14383–14388.
25. Weil T, Korb J, Rehli M (2009) Comparison of queen-specific gene expression in related lower termite species. *Mol Biol Evol* 26:1841–1850.
26. Korb J, Weil T, Hoffmann K, Foster KR, Rehli M (2009) A gene necessary for reproductive suppression in termites. *Science* 324:758.
27. Gotzek D, Ross KG (2009) Current status of a model system: The gene *Gp-9* and its association with social organization in fire ants. *PLoS ONE* 4:e7713.
28. Keller L, Ross KG (1998) Selfish genes: A green beard in the red fire ant. *Nature* 394: 573–575.

29. Krieger MJB, Ross KG (2005) Molecular evolutionary analyses of the odorant-binding protein gene *Gp-9* in fire ants and other *Solenopsis* species. *Mol Biol Evol* 22: 2090–2103.
30. Gotzdek D, Shoemaker DD, Ross KG (2007) Molecular variation at a candidate gene implicated in the regulation of fire ant social behavior. *PLoS ONE* 2:e1088.
31. Leal WS, Ishida Y (2008) *GP-9s* are ubiquitous proteins unlikely involved in olfactory mediation of social organization in the red imported fire ant, *Solenopsis invicta*. *PLoS ONE* 3:e3762.
32. Silva AJ, Kogan JH, Frankland PW, Kida S (1998) CREB and memory. *Annu Rev Neurosci* 21:127–148.
33. Viljakainen L, Pamilo P (2008) Selection on an antimicrobial peptide *defensin* in ants. *J Mol Evol* 67:643–652.
34. Bulmer MS, Crozier RH (2004) Duplication and diversifying selection among termite antifungal peptides. *Mol Biol Evol* 21:2256–2264.
35. Bulmer MS, Lay F, Hamilton C (2010) Adaptive evolution in subterranean termite antifungal peptides. *Insect Mol Biol* 19:669–674.
36. Bulmer MS, Crozier RH (2006) Variation in positive selection in termite GNBPs and Relish. *Mol Biol Evol* 23:317–326.
37. Siomi MC, Mannen T, Siomi H (2010) How does the royal family of *Tudor* rule the PIWI-interacting RNA pathway? *Genes Dev* 24:636–646.
38. Beye M, Hasselmann M, Fondrk MK, Page RE, Omholt SW (2003) The gene *csd* is the primary signal for sexual development in the honeybee and encodes an SR-type protein. *Cell* 114:419–429.
39. Hasselmann M, et al. (2008) Evidence for convergent nucleotide evolution and high allelic turnover rates at the complementary sex determiner gene of Western and Asian honeybees. *Mol Biol Evol* 25:696–708.
40. Hasselmann M, et al. (2008) Evidence for the evolutionary nascence of a novel sex determination pathway in honeybees. *Nature* 454:519–522.
41. Drapeau MD, Albert S, Kucharski R, Prusko C, Maleszka R (2006) Evolution of the Yellow/Major Royal Jelly Protein family and the emergence of social behavior in honey bees. *Genome Res* 16:1385–1394.
42. Zhou X, Tarver MR, Scharf ME (2007) Hexamerin-based regulation of juvenile hormone-dependent gene expression underlies phenotypic plasticity in a social insect. *Development* 134:601–610.
43. Zhou X, Tarver MR, Bennett GW, Oi FM, Scharf ME (2006) Two hexamerin genes from the termite *Reticulitermes flavipes*: Sequence, expression, and proposed functions in caste regulation. *Gene* 376:47–58.
44. Kunieda T, et al. (2006) Carbohydrate metabolism genes and pathways in insects: Insights from the honey bee genome. *Insect Mol Biol* 15:563–576.
45. Le Conte Y, Hefetz A (2008) Primer pheromones in social hymenoptera. *Annu Rev Entomol* 53:523–542.
46. Enard W, et al. (2002) Molecular evolution of *FOXP2*, a gene involved in speech and language. *Nature* 418:869–872.
47. Wagner GP, Pavlicev M, Cheverud JM (2007) The road to modularity. *Nat Rev Genet* 8:921–931.
48. Robertson HM, Wanner KW (2006) The chemoreceptor superfamily in the honey bee, *Apis mellifera*: Expansion of the odorant, but not gustatory, receptor family. *Genome Res* 16:1395–1403.
49. Engsontia P, et al. (2008) The red flour beetle's large nose: An expanded odorant receptor gene family in *Tribolium castaneum*. *Insect Biochem Mol Biol* 38:387–397.
50. Robertson HM, Gadau J, Wanner KW (2010) The insect chemoreceptor superfamily of the parasitoid jewel wasp *Nasonia vitripennis*. *Insect Mol Biol* 19(Suppl1):121–136.
51. Nei M, Niimura Y, Nozawa M (2008) The evolution of animal chemosensory receptor gene repertoires: Roles of chance and necessity. *Nat Rev Genet* 9:951–963.
52. Plettner E, et al. (1997) Species- and caste-determined mandibular gland signals in honeybees (*Apis*). *J Chem Ecol* 23:363–377.
53. Urbanová K, Cahliková L, Hovorka O, Ptáček V, Valterová I (2008) Age-dependent changes in the chemistry of exocrine glands of *Bombus terrestris* queens. *J Chem Ecol* 34:458–466.
54. Cruz-López L, et al. (2005) Mandibular gland secretion of *Melipona beecheii*: chemistry and behavior. *J Chem Ecol* 31:1621–1632.
55. Tokuda G, Saito H, Watanabe H (2002) A digestive β -glucosidase from the salivary glands of the termite, *Neotermes koshunensis* (Shiraki): Distribution, characterization and isolation of its precursor cDNA by 5'- and 3'-RACE amplifications with degenerate primers. *Insect Biochem Mol Biol* 32:1681–1689.
56. Dunbar RIM, Shultz S (2007) Evolution in the social brain. *Science* 317:1344–1347.
57. Oster GF, Wilson EO (1979) *Caste and Ecology in the Social Insects* (Princeton Univ Press, Princeton).
58. Gronenberg W, RIVERS AJ (2009) *Organization of Insect Societies: From Genome to Sociocomplexity*, eds Gadau J, Fewell J (Harvard Univ Press, Cambridge, MA), pp 377–401.
59. O'Donnell S, Donlan N, Jones T (2007) Developmental and dominance-associated differences in mushroom body structure in the paper wasp *Mischocyttarus mastigophorus*. *Dev Neurobiol* 67:39–46.
60. Salvador A, Costa R (2009) Coping with competition: Neuroendocrine responses and cognitive variables. *Neurosci Biobehav Rev* 33:160–170.
61. Molina Y, Harris RM, O'Donnell S (2009) Brain organization mirrors caste differences, colony founding and nest architecture in paper wasps (Hymenoptera: Vespidae). *Proc Biol Sci* 276:3345–3351.
62. Wilson-Rich N, Spivak M, Fefferman NH, Starks PT (2009) Genetic, individual, and group facilitation of disease resistance in insect societies. *Annu Rev Entomol* 54: 405–423.
63. Tarpy DR, Seeley TD (2006) Lower disease infections in honeybee (*Apis mellifera*) colonies headed by polyandrous vs monandrous queens. *Naturwissenschaften* 93: 195–199.
64. Lazzaro BP (2008) Natural selection on the *Drosophila* antimicrobial immune system. *Curr Opin Microbiol* 11:284–289.
65. Hoffmann JA (2003) The immune response of *Drosophila*. *Nature* 426:33–38.
66. Evans JD, et al. (2006) Immune pathways and defence mechanisms in honey bees *Apis mellifera*. *Insect Mol Biol* 15:645–656.
67. International Aphid Genomics Consortium (2010) Genome sequence of the pea aphid *Acyrthosiphon pisum*. *PLoS Biol* 8:e1000313.
68. Viljakainen L, et al. (2009) Rapid evolution of immune proteins in social insects. *Mol Biol Evol* 26:1791–1801.
69. Stoven S, et al. (2003) Caspase-mediated processing of the *Drosophila* NF-kappaB factor Relish. *Proc Natl Acad Sci USA* 100:5991–5996.
70. Liao Z, Jia Q, Li F, Han Z (2010) Identification of two piwi genes and their expression profile in honeybee, *Apis mellifera*. *Arch Insect Biochem Physiol* 74:91–102.
71. Cook JM (1993) Sex determination in the Hymenoptera: A review of models and evidence. *Heredity* 71:421–435.
72. Kamakura M (2011) Royalactin induces queen differentiation in honeybees. *Nature*, 10.1038/nature10093.
73. Lyko F, et al. (2010) The honey bee epigenomes: Differential methylation of brain DNA in queens and workers. *PLoS Biol* 8:e1000506.
74. Yu F, Mao F, Jianke L (2010) Royal jelly proteome comparison between *A. mellifera* ligustica and *A. cerana cerana*. *J Proteome Res* 9:2207–2215.
75. Ferguson LC, Green J, Surridge A, Jiggins CD (2011) Evolution of the insect yellow gene family. *Mol Biol Evol* 28:257–272.
76. Webster T, Peng Y (1988) The evolution of food-producing glands in eusocial bees (Apoidea, Hymenoptera). *J Evol Biol* 2:165–176.
77. Riddiford LM (2008) Juvenile hormone action: A 2007 perspective. *J Insect Physiol* 54: 895–901.
78. Martins JR, Nunes FMF, Cristino AS, Simões ZLP, Bitondi MMG (2010) The four hexamerin genes in the honey bee: Structure, molecular evolution and function deduced from expression patterns in queens, workers and drones. *BMC Mol Biol* 11:23.
79. Hunt JH, et al. (2010) Differential gene expression and protein abundance evince ontogenetic bias toward castes in a primitively eusocial wasp. *PLoS ONE* 5:e10674.
80. Wheeler DE, Buck NA (1995) Storage proteins in ants during development and colony founding. *J Insect Physiol* 41:885–894.
81. Robinson GE, Vargo EL (1997) Juvenile hormone in adult eusocial Hymenoptera: Gonadotropin and behavioral pacemaker. *Arch Insect Biochem Physiol* 35:559–583.
82. Ament SA, Corona M, Pollock HS, Robinson GE (2008) Insulin signaling is involved in the regulation of worker division of labor in honey bee colonies. *Proc Natl Acad Sci USA* 105:4226–4231.
83. Avndam GV, Norberg K, Hagen A, Omholt SW (2003) Social exploitation of vitellogenin. *Proc Natl Acad Sci USA* 100:1799–1802.
84. Corona M, et al. (2007) Vitellogenin, juvenile hormone, insulin signaling, and queen honey bee longevity. *Proc Natl Acad Sci USA* 104:7128–7133.
85. Zera AJ, Harshman LG, Williams TD (2007) Evolutionary endocrinology: The developing synthesis between endocrinology and evolutionary genetics. *Annu Rev Ecol Evol Syst* 38:793–817.
86. Grönke S, Clarke DF, Broughton S, Andrews TD, Partridge L (2010) Molecular evolution and functional characterization of *Drosophila* insulin-like peptides. *PLoS Genet* 6:e1000857.
87. Alvarez-Ponce D, Aguadé M, Rozas J (2009) Network-level molecular evolutionary analysis of the insulin/TOR signal transduction pathway across 12 *Drosophila* genomes. *Genome Res* 19:234–242.
88. Sappington TW, Raikhel AS (1998) Molecular characteristics of insect vitellogenins and vitellogenin receptors. *Insect Biochem Mol Biol* 28:277–300.
89. Hunt BG, et al. (2010) Sociality is linked to rates of protein evolution in a highly social insect. *Mol Biol Evol* 27:497–500.
90. Suarez RK, et al. (2005) Energy metabolism in orchid bee flight muscles: Carbohydrate fuels all. *J Exp Biol* 208:3573–3579.
91. Roubik DW (1992) *Ecology and Natural History of Tropical Bees* (Cambridge Univ Press, Cambridge, United Kingdom).
92. Southwick EE, Heldmaier G (1987) Temperature control in honey bee colonies. *Bioscience* 37:395–399.
93. Finkel T, Holbrook NJ (2000) Oxidants, oxidative stress and the biology of ageing. *Nature* 408:239–247.
94. Toth AL, Robinson GE (2009) Evo-devo and the evolution of social behavior: Brain gene expression analyses in social insects. *Cold Spring Harb Symp Quant Biol* 74:419–426.
95. Toth AL, et al. (2010) Brain transcriptomic analysis in paper wasps identifies genes associated with behaviour across social insect lineages. *Proc Biol Sci* 277:2139–2148.
96. Nonacs P, Kapheim KM (2008) Social heterosis and the maintenance of genetic diversity at the genome level. *J Evol Biol* 20:2253–2265.
97. Linksvayer TA, Wade MJ (2009) Genes with social effects are expected to harbor more sequence variation within and between species. *Evolution* 63:1685–1696.
98. Johnson BR, Linksvayer TA (2010) Deconstructing the superorganism: Social physiology, groundplans, and sociogenomics. *Q Rev Biol* 85:57–79.
99. Dean AM, Thornton JW (2007) Mechanistic approaches to the study of evolution: The functional synthesis. *Nat Rev Genet* 8:675–688.
100. Feder ME, Mitchell-Olds T (2003) Evolutionary and ecological functional genomics. *Nat Rev Genet* 4:651–657.
101. Charnov EL (1978) Evolution of eusocial behavior: Offspring choice or parental parasitism? *J Theor Biol* 75:451–465.
102. Hughes WOH, Oldroyd BP, Beekman M, Ratnieks FLW (2008) Ancestral monogamy shows kin selection is key to the evolution of eusociality. *Science* 320:1213–1216.